

# Total RNA extraction from *Ulva linza* Linnaeus samples using Ribospin™ Plant kit

## Experimental Conditions

### Materials Required

- ♦ Ribospin™ Plant kit (307-150, 50 preps)
- ♦ Tissuelyser II (85300, Q) (or another bead beating device)
- ♦ Liquid nitrogen (LN<sub>2</sub>)
- ♦ Absolute ethanol (C<sub>2</sub>H<sub>6</sub>O, CAS No. : 64-17-5, ≥99.0%)
- ♦ 70% ethanol (C<sub>2</sub>H<sub>6</sub>O, CAS No. : 64-17-5, 70.0%)
- ♦ 1.5 ml or 2.0 ml microcentrifuge tube
- ♦ Vortex mixer
- ♦ Centrifuge (Max. speed 14,000 rpm)
- ♦ Pipette & sterile pipette tips
- ♦ Suitable protector (ex. lab coat, disposable gloves, goggles, etc.)
- ♦ Ice (for incubation)

### Sample Information

- ♦ Sample type :



*Ulva linza* Linnaeus

- ♦ Sampling : After collecting fresh *Ulva linza* Linnaeus samples, put them in a bag and seal it.
- ♦ How to store : Store in -70°C deep freezer
- ♦ Homogenizing : Tissuelyser II (85300, Supplier : Q)
- ♦ Conditions :
  - Sample amount : 25, 50 and 100 mg
  - Elution volume : 50 µl

## Protocol

### Before experiment

1. Before using for the first time, add absolute ethanol (ACS grade or better) into Buffer RBW and RNW as indicated on the bottle.
2. Prepare DNase I reaction mixture just before step 8.
  - Prepare aliquot DNase I and thaw on ice.
  - Mix 2 µl DNase I with 70 µl Buffer DRB.

### Sample preparation

1. After measuring 25, 50 and 100 mg of each sample, put it into a 1.5 ml microcentrifuge tube. Then, it is rapidly frozen using LN<sub>2</sub>.
2. Grind the sample using Tissuelyser II (30 Hz, 10 sec).
3. The next step is according to the protocol of Ribospin™ Plant.

### Ribospin™ Plant brief protocol

\* For more details, refer to the handbook of Ribospin™ Plant

1. Transfer the powder into a 1.5 ml microcentrifuge tube.
2. Add 350 µl of Buffer RPL and incubate the mixture for 3 min at room temperature.
3. Transfer the lysate to EzPure™ Filter and centrifuge at ≥10,000 x g for 30 sec.
4. Transfer the supernatant into a 1.5 ml microcentrifuge tube.
5. Add 1 volume of 70% ethanol to the supernatant and mix well.
6. Apply the mixture into Column Type W and centrifuge at ≥10,000 x g for 30 sec.
7. Add 500 µl of Buffer RBW to Column Type W and centrifuge at ≥10,000 x g for 30 sec.
8. Apply the DNase I mixture into Column Type W.
9. Incubate the mixture for 10 min at room temperature.
10. Add 500 µl of Buffer RBW to Column Type W and incubate for 2 min and centrifuge at ≥10,000 x g for 30 sec.
11. Add 500 µl of Buffer RNW to Column Type W and centrifuge at ≥10,000 x g for 30 sec (twice).
12. Centrifuge at ≥10,000 x g for an additional 1 min.
13. Add 50 µl of Nuclease-free water to the center of the membrane.
14. Centrifuge at ≥10,000 x g for 1 min.

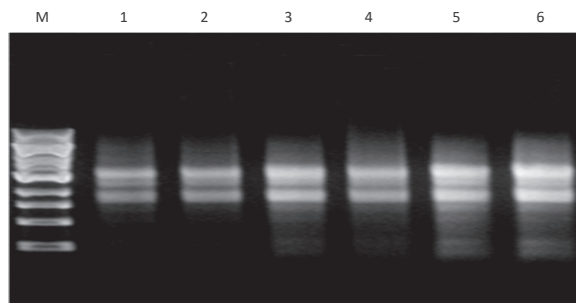
## Result

#	Sample	Conc. (ng/µl)	A <sub>260/280</sub>	A <sub>260/230</sub>	Yield (µg)
1	<i>Ulva linza</i> Linnaeus (25 mg)	100.2	2.19	2.33	5.0
2		111.3	2.24	2.45	5.6
3	<i>Ulva linza</i> Linnaeus (50 mg)	187.7	2.21	2.24	9.4
4		183.6	2.25	2.42	9.2
5	<i>Ulva linza</i> Linnaeus (100 mg)	256.6	2.24	2.44	12.8
6		334.2	2.25	2.46	16.7

Table 1. The concentration, yield and and purity of RNA extracted from 25~100 mg of *Ulva linza* Linnaeus samples.

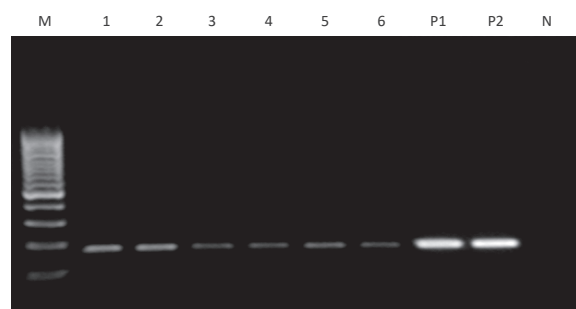
※ Remark  
Absorbance measurement instrument : NanoDrop™ 2000/2000 c (ND-2000, Supplier : T)

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**Figure 1. The result of electrophoresis of total RNA from *Ulva linza* Linnaeus sample.**  
 Lanes 1~2 : 2  $\mu$ l of eluate from 25 mg of *Ulva linza* Linnaeus  
 Lanes 3~4 : 2  $\mu$ l of eluate from 50 mg of *Ulva linza* Linnaeus  
 Lanes 5~6 : 2  $\mu$ l of eluate from 100 mg of *Ulva linza* Linnaeus  
 Lane M : GENESTA™ 1 kb DNA ladder with 5X loading dye (GA-100, GeneAll®, 1  $\mu$ l loading)

※ Remark  
 Electrophoresis conditions : 1.0% agarose gel (150 V, 17 min, 2  $\mu$ l loading)



**Figure 2. The result of electrophoresis of PCR products after RT-PCR.**  
 Lanes 1~2 : 2  $\mu$ l of eluate from 25 mg of *Ulva linza* Linnaeus  
 Lanes 3~4 : 2  $\mu$ l of eluate from 50 mg of *Ulva linza* Linnaeus  
 Lanes 5~6 : 2  $\mu$ l of eluate from 100 mg of *Ulva linza* Linnaeus  
 Lane M : GENESTA™ 100 bp DNA ladder with 5X loading dye (GA-010, GeneAll®, 1  $\mu$ l loading)  
 Lanes P1~P2 : Positive control (total RNA template from pine leaf)  
 Lane N : Negative control (nuclease-free water)

※ Remark  
 Primer : NAD5 gene primer (gene of mitochondrial plant cell)  
 Electrophoresis condition : 2.0% agarose gel (150 V, 20 min, 3  $\mu$ l loading)

## Conclusion

The *Ulva Linnaeus* is well known for the wide distribution throughout the world and the research about its genomic information is widely conducted owe to its major contribution to biofouling.

The isolation of high-quality RNA from *Ulva Linnaeus* is the initial but the most import step for the successful research but there has always been problematic due to a high content a polyphenolic compounds and polysaccharides in the sample.

This study presents simple and effective method for the high-quality RNA isolation from *Ulva Linnaeus* in different starting amounts using Ribospin™ Plant Kit.

The RNA obtained from *Ulva Linnaeus* was found to be suitable for many downstream applications such as RT-PCR, PCR, qPCR, and etc.